



Instruction for Use

***ApexHFHS* DNA Polymerase FS**

AG12201

Version.V3E1

**Research Use Only
Not For Diagnosis Procedures**

1. Description

ApexHF HS DNA Polymerase FS is a high-fidelity DNA polymerase featuring high amplification efficiency, high sensitivity, high specificity, efficient annealing, and rapid extension speed. The optimized PCR reaction system in this product incorporates a unique extension factor, enhancing annealing efficiency and extension speed, significantly shortening the extension time and enabling fast PCR reactions. Additionally, this product contains a monoclonal antibody that inhibits DNA polymerase activity at room temperature, allowing Hot Start PCR. This effectively prevents primer dimer formation and non-specific amplification, ensuring superior PCR performance.

2. Kit Information

Kit Name	Cat. No	Specification
<i>ApexHF</i> HS DNA Polymerase FS	AG12201	50 rxns / 50 μ l

3. Transportation and Storage

Storage	Store at -20°C
Transportation	Transport at -20°C Dry Ice or Blue Ice Condition

4. Kit Components

Cat. No AG 12201

Kit Components	Volume
<i>ApexHF</i> FS DNA Polymerase FS (1U/ μ l)	50 μ l
5X <i>ApexHF</i> FS Buffer (Mg^{2+} plus)*	500 μ l
dNTP Mix (10 mM each)	50 μ l

* Mg^{2+} concentration is 5 mM

** At 74°C for 30 minutes, using activated salmon sperm DNA as the template/primer, the amount of enzyme required to incorporate 10 nmol of deoxynucleotides into acid-insoluble material is defined as one unit of enzyme activity (U).

5. Protocol

5.1 Reagent Preparation

The final reaction volume in this protocol is 50 μ l. The volumes given here may be scaled for larger or smaller reaction volume.

Components	Final Concentration	Volume
<i>ApexHFHS</i> DNA Polymerase FS (1U/ μ l)	1U	1 μ l
5X <i>ApexHFHS</i> Buffer (Mg ²⁺ plus)	1X	10 μ l
dNTP Mix (10 mM each)	0.2 mM	1 μ l
Primer F (10 μ M) *2	0.2 μ M	1 μ l
Primer R (10 μ M) *2	0.2 μ M	1 μ l
Template	\leq 200 ng*3	-
RNase free water	-	Up to 50 μ l

*1: Thaw the reagent tube, and vigorously vortex for 30–60 sec to ensure homogeneity before use. Briefly centrifuge to collect contents at the bottom of the tube.

*2: Recommended final concentration for Primer is 0.2 μ M, could be optimized between 0.1 ~ 0.4 μ M.

*3: Recommended final concentration is less than 200 ng. For template more than 200 ng, PCR program extension step could be adjusted between 10 - 60 sec /kb.

5.2 Thermal Cycling Program

The cycling parameters below are offered as a guideline and may be modified as necessary for optimal results.

Program for 3 Step PCR

Step	Temperature	Time	Number of Cycles
Initial-Denaturation*1	94°C	30 sec	1
Denaturation*2	98°C	10 sec	
Annealing*3	55°C	5 or 15 sec	25~35
Extension	72°C	5 sec / kb*4	

*1: For simple structure template, Initial-Denaturation could be skipped;

For complex template, recommended Initial-Denaturation setup is 94°C for 30 sec-1min.

*2: Could be adjusted per instrument. Recommended to be 94°C for 10-15 sec, or 98°C for 5-10 sec.

*3: Annealing Temperature is recommended to be 5 or 15 sec to prevent smear of electrophoresis result.

When T_m value > 55°C, annealing time shall be 5 sec; for T_m value < 55°C, annealing time shall be 15 sec.

*4: Recommended extension setup is 5 sec/kb; extension time could be extended to 10-60 sec/kb for complex templates or unsatisfying trials.

*5: 2 Step PCR program (as below) could be adopted for high T_m values or unsatisfying result using 3 Step PCR.

* Program for 2 Step PCR

Step	Temperature	Time	Number of Cycles
Initial-Denaturation	94°C	30 sec	1
Denaturation	98°C	10 sec	
Extension	68°C	5 sec / kb	25~35

6. Result Analysis

Purified PCR product could be analyzed via Agarose Gel Electrophoresis.



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