



Instruction for Use

***ApexHFHS* DNA Polymerase FS Master Mix (Dye Plus)**

AG12206

Version.V3E1

**Research Use Only
Not For Diagnosis Procedures**

1. Description

This product is a 2× concentration ready-to-use PCR reaction premix of ApexHF HS DNA Polymerase FS. When performing the PCR reaction, simply add template, primers, and water to the premix for amplification. Additionally, this product contains a dye reagent (light purple-red) required for electrophoresis detection. After the PCR reaction is complete, the sample can be directly subjected to gel electrophoresis, and the reaction mixture will appear purple-red. This premix scheme is easy to operate, minimizes human error to the greatest extent, and allows for rapid detection results. This product features high amplification efficiency, high sensitivity, high specificity, high annealing efficiency, and fast extension speed, enabling rapid PCR reactions. Moreover, the product also contains a monoclonal antibody that can inhibit DNA polymerase activity at room temperature, allowing for Hot Start PCR. This effectively suppresses primer dimer formation and non-specific amplification.

2. Kit Information

Kit Name	Cat. No	Specification
<i>ApexHF</i> HS DNA Polymerase FS Master Mix (Dye Plus)	AG12206	100 rxns / 50 µl

3. Transportation and Storage

Storage	Store at -20°C
Transportation	Transport at -20°C Dry Ice or Blue Ice Condition

4. Kit Components

Cat. No AG 12206

Kit Components	Volume
2X <i>ApexHF</i> FS PCR Master Mix (Dye Plus)*	500 µl x 5 pcs

* Mg²⁺ concentration is 2 mM, dNTPs concentration is 400 µM.

5. Protocol

5.1 Reagent Preparation

The final reaction volume in this protocol is 50 μ l. The volumes given here may be scaled for larger or smaller reaction volume.

Components	Final Concentration	Volume
2X <i>ApexHFFS</i> PCR Master Mix (dye plus) ^{*1}	1 X	25 μ l
Primer F (10 μ M)*2	0.2 μ M	1 μ l
Primer F (10 μ M)*2	0.2 μ M	1 μ l
Template	\leq 200 ng ^{*3}	-
RNase free water	-	Up to 50 μ l

*1: Thaw the reagent tube, and vigorously vortex for 30–60 sec to ensure homogeneity before use. Briefly centrifuge to collect contents at the bottom of the tube.

*2: Recommended final concentration for Primer is 0.2 μ M, could be optimized between 0.1 ~ 0.4 μ M.

*3: Recommended final concentration is less than 200 ng. For template more than 200 ng, PCR program extension step could be adjusted between 10 - 60 sec /kb.

5.2 Thermal Cycling Program

The cycling parameters below are offered as a guideline and may be modified as necessary for optimal results.

Program for 3 Step PCR

Step	Temperature	Time	Number of Cycles
Initial-Denaturation ^{*1}	94°C	30 sec	1
Denaturation ^{*2}	98°C	10 sec	
Annealing ^{*3}	55°C	5 or 15 sec	25~35
Extension	72°C	5 sec / kb ^{*4}	

*1: For simple structure template, Initial-Denaturation could be skipped;

For complex template, recommended Initial-Denaturation setup is 94°C for 30 sec-1min.

*2: Could be adjusted per instrument. Recommended to be 94°C for 10-15 sec, or 98°C for 5-10 sec.

*3: Annealing Temperature is recommended to be 5 or 15 sec to prevent smear of electrophoresis result.

When T_m value > 55°C, annealing time shall be 5 secs; for T_m value < 55°C, annealing time shall be 15 secs.

*4: Recommended extension setup is 5 sec/kb; extension time could be extended to 10-60 sec/kb for complex templates or unsatisfying trials.

*5: 2 Step PCR program (as below) could be adopted for high T_m values or unsatisfying result using 3 Step PCR.

*5 Program for 2 Step PCR

Step	Temperature	Time	Number of Cycles
Initial-Denaturation	94°C	30 sec	1
Denaturation	98°C	10 sec	
Extension	68°C	5 sec / kb	25~35

6. Result Analysis

Purified PCR product could be analyzed via Agarose Gel Electrophoresis.



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